

# Relative Oral Bioavailability Study of Liposomal Curcumin

Dr. Yogesh Dound <sup>1</sup>, Dr. Ramesh Jayaraman <sup>2</sup>

## Abstract

**Background:** Curcumin is a naturally derived yellow polyphenolic compound from the rhizome *Curcuma longa*. Curcumin modulates various signalling pathways, such as Cyclooxygenase-2 (COX-2), Matrix metalloproteinases (MMPs), glutathione, protein kinase C, ATPase, nuclear Nuclear Factor kappa-light-chain-enhancer of activated B cells (NF- $\kappa$ B), Activator protein 1 (AP-1), P-glycoprotein 1 (P-gp), Multidrug resistance-associated proteins (MRP-1, MRP-2), Receptor tyrosine-protein kinase (ErbB2), Alpha-1-acid glycoprotein (AGP), Cyclin D1 etc. Curcumin, as a free drug, has a short plasma half-life because it is rapidly metabolised in the liver. Its oral bioavailability is extremely poor, and hence Curcumin is classified as a class IV compound in the BCS system.

**Materials and Methods:** Authors of the current study have formulated Liposomal Curcumin and conducted a Relative Oral Bioavailability Study of Liposomal Curcumin in comparison with plain Curcumin in rats. The treatment group was given Curcumin 25% lipo at the dose of 500 mg/kg body weight and the control group was given plain Curcumin 500 mg/kg body weight of the rats.

**Observations:** Oral administration of Liposomal Curcumin showed rapid absorption ( $t_{max}$  = 15 min) with Peak plasma concentration ( $C_{max}$ ), Area under Curve (AUC) and half-life ( $t_{1/2}$ ) of 42.3 ng/ml, 244 ng.h/ml and 5.5 h, respectively. In the control group, the Curcumin levels were below the level of quantitation (5.43 ng/ml).

**Conclusion:** This study shows that Liposomal Curcumin has higher bioavailability as compared to the marketed formulations of Curcumin. Further extensive clinical studies are needed to prove efficacy of Liposomal Curcumin in various human applications.

**Keywords:** Liposomal Curcumin, bioavailability, pharmacokinetics

**Conflict of Interest:** Dr. Yogesh Dound is Proprietor, Shreepad Shree Vallabh SSV Phytopharmaceuticals.

**Source of funding:** Shreepad Shree Vallabh SSV Phytopharmaceuticals funded the study.

<sup>1</sup> Proprietor, Shreepad Shree Vallabh SSV Phytopharmaceuticals, Mumbai

<sup>2</sup> Chief Scientific Officer, TheraIndx LifeSciences, Pvt. Ltd., Bengaluru

**Corresponding author:** Dr. Yogesh Dound, Proprietor, Shreepad Shree Vallabh SSV Phytopharmaceuticals, Mumbai.

Email: yogesh\_dound@yahoo.com

## Introduction

Curcumin is a naturally derived yellow polyphenolic compound from the rhizome *Curcuma longa*.<sup>[1]</sup> Curcumin, as a free drug, has a short plasma half-life because it is rapidly metabolised in the liver.<sup>[2]</sup> The wide range of activities of Curcumin are confirmed to its ability to modulate various signalling pathways, such as Cyclooxygenase-2 (COX-2), Matrix metalloproteinases (MMPs), glutathione, protein kinase C, ATPase, nuclear Nuclear Factor kappa-light-chain-enhancer of activated B cells (NF- $\kappa$ b), Activator protein 1 (AP-1), P-glycoprotein 1 (P-gp), Multidrug resistance-associated proteins (MRP-1, MRP-2), Receptor tyrosine-protein kinase (ErbB2), Alpha-1-acid glycoprotein AGP), Cyclin D1, etc.<sup>[3,4]</sup> Various pre-clinical and clinical studies have confirmed that Curcumin is significantly non-toxic even at very high doses.<sup>[5]</sup> Unfortunately, Curcumin faces different problems for its application, such as low-aqueous solubility, rapid systemic clearance, inadequate tissue absorption and degradation at alkaline pH values. Also, Curcumin is rapidly photodegraded by light, hence limiting its clinical use.<sup>[6,7]</sup> Consequently, its oral bioavailability is extremely poor, and Curcumin is classified as a class IV compound in the BCS system.<sup>[8]</sup>

To overcome these limitations, various strategies such as liposomes, solid dispersion, complex, emulsion, micelles, nanogels and microspheres have been employed to overcome poor absorption and other pharmacokinetic limitations of curcumin. Further, Curcumin is poorly water-soluble, <0.125 mg/L, and susceptible to enzymatic degradation in the blood stream, all of which limit the drug's bioavailability and therapeutic potential. To improve the bioavailability of curcumin, many different approaches, have been developed for sustained and efficient curcumin delivery.<sup>[9]</sup>

## Materials

Curcumin 25% I lipo (Liposomal Curcumin) was obtained from Shreepad Shree Vallabh SSV Phytopharmaceuticals, Mumbai. Male Wistar rats (290–340 g) were housed in a temperature (23  $\pm$  1°C) and light-controlled room (12 hours light/dark cycle). They were allowed ad libitum access to food and water for 7 days. Rats were randomly divided into two groups of eight animals each. The control group was given plain Curcumin 500 mg/kg body weight of the rats and the treatment group was given Curcumin 25% I lipo at the dose of 500 mg/kg body weight. The study was performed as per ethical practices laid down in

the CPCSEA guidelines<sup>[10]</sup> for animal care and use. The study was approved by the Institutional Animals Ethics Committee (IAEC) of the test facility.

## Method

All the formulations were orally administered by oral gavage by means of flexible plastic tubes with 5 cm in length. At pre-determined interval (0, 5, 15, 30, 60, 120, 180, 240 and 480 min) after administration, blood was harvested from the abdominal Vena Cava of rats under light anesthesia (di ethyl ether) and placed into heparinized tubes to prevent clotting. Plasma was immediately prepared by centrifugation at 1000  $\times$  g for 15 min at 4°C and stored at -80°C until analysis. Plasma (200 mL) was mixed with 200 mL of 0.1 M sodium phosphate buffer (pH6.8) containing 0.1% EDTA, 200 mL of distilled water, and 600 mL of methanol. The mixture was vortexed for 3 min and, after the addition of 4mL of hexane, was shaken vigorously and centrifuged at 1000  $\times$  g for 10 min at 4°C. The hexane layer was discarded, and 1.2 mL of distilled water and 3 mL of ethyl acetate were added to the mixture (aqueous-methanol layer). This was shaken vigorously and centrifuged at 1000  $\times$  g for 15 min at 4°C, and the ethyl acetate layer was finally collected. This ethyl acetate extraction was repeated four times. The combined extraction phases were evaporated to dryness in vacuo and the residue was dissolved in 100 mL of methanol and was analysed for Curcumin content by HPLC method.

## Results

Following oral administration of Curcumin 25% I lipo, Liposomal Curcumin showed rapid absorption ( $T_{max}$  =15 min) with Peak plasma concentration ( $C_{max}$ ), Area under Curve (AUC) and half-life ( $t_{1/2}$ ) of 42.3 ng/ml, 244 ng.h/ml and 5.5 h, respectively. In the control group, the Curcumin levels were below the level of quantitation (5.43 ng/ml). The Bioanalytical method was linear between 5.43 ng/ml and 3528.57 ng/ml and met all acceptance criteria. Table 1 shows the average values of the pharmacokinetic parameters in the test group and the control group. Curcumin plasma concentration versus time profiles in both the groups can be seen in Figure 1. Table 2 shows a comparison of the various bioavailability studies on different formulations<sup>[11-15]</sup> of Curcumin following oral administration in rats.

Poly(lactic-co-glycolic acid) (PLGA) and PLGA-poly(ethylene glycol) (PEG) (PLGA-PEG) blend nanoparticles are considered as potential carriers for the oral delivery of curcumin. The nanoparticles pro-

longed extensively the curcumin release. In an *in vivo* pharmacokinetic study, all parameters including mean half-life and the  $C_{max}$  were improved by curcumin nanoparticles. Compared to the curcumin aqueous suspension, the PLGA and PLGA-PEG nanoparticles increased the curcumin bioavailability by 15.6- and 55.4-fold, respectively [11].

$K_e$  in rats [12]. Curcumin incorporated into TMC-coated liposomes exhibited bioavailability ( $C_{max} = 46.13$  ng/ml,  $T_{max} = 2.0 \pm 0.62$  hours), compared with curcumin encapsulated by uncoated liposomes ( $C_{max} = 32.12$  ng/ml,  $T_{max} = 2.0 \pm 0.30$  hours) and curcumin suspension ( $C_{max} = 35.46$  ng/ml,  $T_{max} = 1.0 \pm 0.22$  hours).

Curcumin powder extracted from Indian turmeric by using alcohol was evaluated *in vivo* for the plasma pharmacokinetics at the dose of 50 mg/kg and 300 mg/kg. For both the doses the  $T_{max}$  was 2 hours and the  $C_{max}$  was  $13.0 \pm 5.8$  and  $37.4 \pm 36.1$  ng/ml respectively. [13] In another study, the Curcumin powder at the dose of 250 mg/kg was evaluated for oral bioavailability studies *in vivo*. [14] The  $C_{max}$  was  $32.29 \pm 14.93$  ng/ml and the

**Table 1. Mean plasma concentration/time curves and mean pharmacokinetic parameters obtained after oral administration to rats in both the groups.**

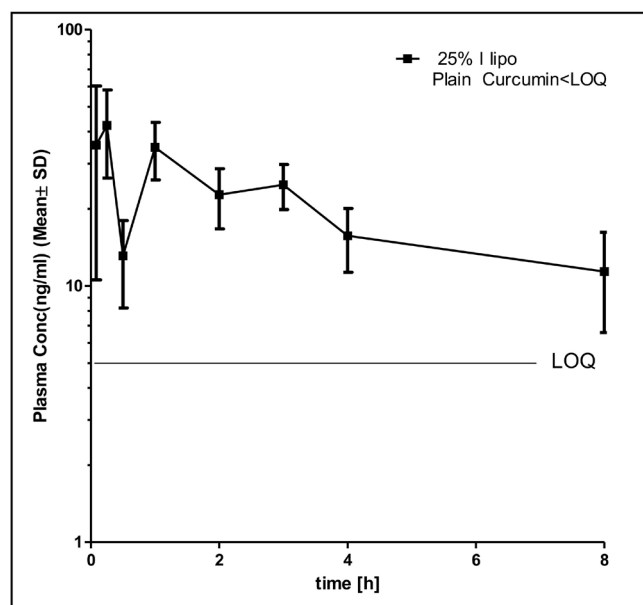
Pharmacokinetic Parameters	Curcumin 25% I lipo	Plain Curcumin
	Estimate	Estimate
$K_e$ (1/h)	0.126	NE
$t_{1/2}$ (h)	5.520	NE
$T_{max}$ (min)	15	NE
$C_{max}$ (ng/ml)	42.280	NE
$T_{last}$ (h)	8.000	NE
$C_{last}$ (ng/ml)	11.388	NE
$AUC_{last}$ (h*ng/ml)	154	NE
$AUC_{INF}$ (h*ng/ml)	244	NE
$AUC_{\%Extrap}$	37	NE

NE: Not Estimated as Curcumin levels were BLOQ [5.43 ng/ml]  
BLOQ: Below the limit of Quantitation

$K_e$  - Elimination rate constant;  $t_{1/2}$  - Elimination half life;  $T_{max}$  - Time of maximum drug concentration;  $C_{max}$  - Maximum plasma concentration;  $T_{last}$  - Time of  $C_{last}$ ;  $C_{last}$  - last observed quantifiable concentration;  $AUC_{last}$  - Area under the plasma concentration-time curve from time zero to time of last measurable concentration;  $AUC_{INF}$  - Area under the plasma concentration-time curve from time zero to infinity;  $AUC_{\%Extrap}$  - Area under the plasma concentration-time curve extrapolated from time t to infinity as a percentage of total AUC.

$T_{max}$  was around 1 hour. In a similar study with crystalline Curcumin powder at the dose of 100 mg/kg the  $C_{max}$  was  $35 \pm 8.0$  ng/ml and the  $T_{max}$  was almost 80 minutes. [15]

Curcumin 25% I lipo when compared with other similar formulations or different formulations has shown improved pharmacokinetic profiles in terms of rapid absorption ( $T_{max}$ ) with Peak plasma concentration ( $C_{max}$ ), Area under Curve (AUC) and half-life ( $t_{1/2}$ ) leading to higher bioavailability in comparison to various Curcumin formulations studied.



**Figure 1. Curcumin plasma concentration versus time profiles in both the groups**

## Discussion

Liposomes are useful vehicles to deliver curcumin because of their ability to improve bioavailability and solubility of curcumin. They can be used as a potential targeted delivery system by folate-ligand surface modification, gradual release of curcumin in the body, and subsequent improved efficacy of treatment in cancer patients. [16,17] Liposomes have been used in the delivery of anticancer drugs and are able to alter the biodistribution and clearance of drug molecules. [18,19] Liposomal nanoconstructs are extensively studied to improve the bioavailability of Curcumin, as these drug delivery systems are approved by the Food and Drug Administration (FDA). [20,21]

Studies have shown that effective liposomal encapsulation of Curcumin can increase its solubility to 500 ug/mL as compared to 0.125 mg/L. The benefits of using a phospholipid bilayer nanoconstruct, include du-

**Table 2. Peak plasma/serum concentration ( $C_{max}$ ) and time to reach peak plasma/serum ( $T_{max}$ ) concentration obtained from various studies with different formulations of Curcumin.**

Curcumin formulation	Curcumin (g/kg)	$C_{max}$ (ng/ml)	$T_{max}$ (hour)	Ref.
Curcumin unformulated	0.05	4.066 ± 0.564	0.50	[11]
PLGA Curcumin nanoparticles	0.05	11.783 ± 0.454	2.00	[11]
PLGA-PEG Curcumin nanoparticles	0.05	29.778 ± 4.632	3.00	[11]
Curcumin suspension	0.25	35.46 ± 12.81	1.00	[12]
TMC-coated Curcumin liposomes	0.04	46.13 ± 5.31	2.00	[12]
Uncoated Curcumin liposomes	0.04	32.12 ± 9.42	2.00	[12]
Curcumin Powder	0.05	13.0 ± 5.8	2.00	[13]
	0.3	37.4 ± 36.1	2.00	[13]
Raw Curcumin	0.25	32.29 ± 14.93	0.58	[14]
Curcumin crystal suspension	0.10	35.00 ± 8.0	1.33	[15]

al loading of hydrophilic and hydrophobic drugs, improved bioavailability to target site cell and tissue, stability of encapsulated drugs, pharmacological inactivity and minimal toxicity of phospholipids. Several *in vitro* studies have shown the benefits of encapsulating curcumin in different cancer types such as pancreatic adenocarcinoma, osteosarcoma, liver cancer etc. [22-25] Liposomal delivery of Curcumin has also been tested in clinical trials for cancer therapy. Storka *et al.* found that short-term administration of intravenous doses up to 120 mg/m<sup>2</sup> are safe in healthy individuals. [26] Several products like Theracurmin® (Made with gum ghatti) is used to improve gut absorption and enhance the effects of curcumin in the body which range from cardiovascular health to free radical protection. [22,23] Other product Longvida® use solid lipid curcumin particle (SLCP) technology to enhance the bioavailability of curcumin by protecting curcumin from degradation in the stomach and promoting free drug absorption in the gut.

In the current study, the pharmacokinetic profile obtained for Liposomal Curcumin was compared with the pharmacokinetic profiles  $C_{max}$  and  $T_{max}$  of various formulations of Curcumin published in literature [11-15] and was found to be superior in comparison.

The results clearly show that Curcumin 25% I lipo (Liposomal Curcumin) is highly bioavailable and this property will help improve its efficacy for various human applications.

## Conclusion

Curcumin 25% I lipo (Liposomal Curcumin) is

highly bioavailable as seen by the pharmacokinetic profiles i.e. peak plasma/serum concentration ( $C_{max}$ ) and time to reach peak plasma/serum ( $T_{max}$ ) concentration. These pharmacokinetic values need to be extrapolated and correlated in humans. For the same, further extensive clinical studies are needed to be conducted. This will also prove the efficacy of Curcumin 25% I lipo (Liposomal Curcumin) in various indications in comparison to currently available formulations of Curcumin.

## References:

1. Miller JM, Thompson JK, MacPherson MB, Beuschel SL, Westbom CM, Sayan M, *et al.* Curcumin: a double hit on malignant mesothelioma. *Cancer Prev Res (Phila)*. 2014; 7: 330-40.
2. Hussain Z, Thu HE, Amjad MW, Hussain F, Ahmed TA, Khan S. Exploring recent developments to improve antioxidant, anti-inflammatory and antimicrobial efficacy of curcumin: A review of new trends and future perspectives. *Mater Sci Eng C Mater Biol Appl*. 2017; 77: 1316-26.
3. Anand, P., Sundaram, C., Jhurani, S., Kunnumakkara, A. B., and Aggarwal, B. B. Curcumin and cancer: an "old-age" disease with an "age-old" solution. *CancerLett*. 2008. 267, 133-164.
4. Kunnumakkara, A.B., Anand, P., and Aggarwal, B.B. Curcumin inhibits proliferation, invasion, angiogenesis and metastasis of different cancers through interaction with multiple cell signalling proteins. *CancerLett*. 2008. 269, 199-225.
5. Mohanty, C., Das, M., and Sahoo, S.K.. Emerging role of nano carriers to Increase the solubility and bioavailability of curcumin. *Expert Opin. DrugDeliv*. 2012. 9, 1347-1364.
6. Cañamares, M. V., Garcia-Ramos, J. V., and Sanchez-Cortes,



- S. Degradation of curcumin dye in aqueous solution and on Ag nanoparticles studied by ultraviolet-visible absorption and surface-enhanced Raman spectroscopy. *Appl.Spectrosc.* 2006. 60, 1386–1391.
7. Kumavat, S. D., Chaudhari, Y. S., Borole, P., Mishra, P., Shenghani, K., and Duvvuri, P. Degradation studies of curcumin. *Int. J.Pharm.Rev.Res* 2016. 3, 50–55.
  8. Wahlang, B., Pawar, Y. B., and Bansal, A. K. (2011). Identification of permeability related hurdles in oral delivery of curcumin using the CaCo-2 cell model. *Eur. J. Pharm. Biopharm.* 77, 275–282.
  9. Liu A, Lou H, Zhao L, Fan P (2006) Validated LC/MS/MS assay for curcumin and tetrahydrocurcumin in rat plasma and application to pharmacokinetic study of phospholipid complex of curcumin. *J Pharm Biomed Anal* 40(3):720–727.
  10. CPCSEA Guidelines for Laboratory Animal Facility. *Indian Journal of Pharmacology* 2003. 35: 257- 274.
  11. N. M. Khalil, T. C. F. do Nascimento, D. M. Casa *et al.*, Pharmacokinetics of curcumin-loaded PLGA and PLGA-PEG blend nanoparticles after oral administration in rats. *Colloids and Surfaces B: Biointerfaces*, 2013. 101, 353–360.
  12. Chen H, Wu J, Sun M, Guo C, Yu A, Cao F *et al.* N-trimethyl chitosan chloride-coated liposomes for the oral delivery of curcumin. *J Liposome Res* 2012;22(2):100-9.
  13. Sasaki H, Sunagawa Y, Takahashi K, Imaizumi A, Fukuda H *et al.* Innovative Preparation of Curcumin for Improved Oral Bioavailability. *Biol Pharm Bull* 2011;34(5):660-5.
  14. Pawar Y, Purohit H, Valicherla GR, Munjal B, Lale S, Patel SB *et al.* Novel lipid based oral formulation of curcumin: Development and optimization by design of experiments approach. *International Journal of Pharmaceutics* 2012; 436: 617–23.
  15. Onoue S, Takahashi H, Kawabata Y, Seto Y, Hatanaka J, Timmermann B *et al.* Formulation Design and Photochemical Studies on Nanocrystal Solid Dispersion of Curcumin with Improved Oral Bioavailability. *Journal of Pharmaceutical Sciences* 2010;99(4): 1871-81.
  16. Feng T, Wei Y, Lee RJ, Zhao L. Liposomal curcumin and its application in cancer. *Int J Nanomedicine.* 2017; 12: 6027-44.
  17. Bisht S, Schlesinger M, Rupp A, Schubert R, Nolting J, Wenzel J, *et al.* A liposomal formulation of the synthetic curcumin analog EF24 (Lipo-EF24) inhibits pancreatic cancer progression: towards future combination therapies. *J Nanobiotechnology.* 2016; 14: 57.
  18. Bingham RJ, Olmsted PD, Smye SW. Undulation instability in a bilayer lipid membrane due to electric field interaction with lipid dipoles. *Phys Rev E Stat Nonlin Soft Matter Phys.* 2010;81(5 pt 1):703–708.
  19. Wang XY, Ishida T, Ichihara M, Kiwada H. Influence of the physicochemical properties of liposomes on the accelerated blood clearance phenomenon in rats. *J Control Release.* 2005;104(1):91–102.
  20. Sun D, Zhuang X, Xiang X, Liu Y, Zhang S, Liu C, *et al.* A novel nanoparticle drug delivery system: the anti-inflammatory activity of curcumin is enhanced when encapsulated in exosomes. *Mol Ther.* 2010; 18: 1606-14.
  21. He W, Zou C, Tian Z, Tan W, Shen W, Chen J, *et al.* Nasopharyngeal carcinoma treated with bevacizumab combined with paclitaxel liposome plus cisplatin: a case report and literature review. *Onco Targets Ther.* 2017; 10:67-72.
  22. Mahmud M, Piwoni A, Filipczak N, Janicka M, Gubernator J. Long-Circulating Curcumin-Loaded Liposome Formulations with High Incorporation Efficiency, Stability and Anticancer Activity towards Pancreatic Adenocarcinoma Cell Lines In Vitro. *PLoS One.* 2016; 11: e0167787.
  23. Kitajima H, Komizu Y, Ichihara H, Goto K, Ueoka R. Hybrid liposomes inhibit tumor growth and lung metastasis of murine osteosarcoma cells. *Cancer Med.* 2013; 2: 267-76.
  24. Ucisik MH, Kupcu S, Schuster B, Sleytr UB. Characterization of CurcuEmulsomes: nanoformulation for enhanced solubility and delivery of curcumin. *J Nanobiotechnology.* 2013; 11: 37.
  25. Yallapu MM, Jaggi M, Chauhan SC. Curcumin nano formulations: A future nanomedicine for cancer. *Drug Discov Today.* 2012; 17: 71-80.
  26. Storka A, Vcelar B, Klickovic U, Gouya G, Weisshaar S, Aschauer S, *et al.* Safety, tolerability and pharmacokinetics of liposomal curcumin in healthy humans. *Int J Clin Pharmacol Ther.* 2015; 53: 54-65.

+